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# Relationship between mitochondrial haplogroup and seasonal changes of physiological responses to cold

Takayuki Nishimura<sup>1\*</sup> and Shigeki Watanuki<sup>2</sup>

## Abstract

**Background:** Physiological responses to cold exhibit individual variation that can be affected by various factors, such as morphological characteristics, seasonal changes, and lifestyle; however, the genetic factors associated with this variation remain unclear. Recent studies have identified mtDNA as a potential genetic factor affecting cold adaptation. In addition, non-shivering thermogenesis (NST), a process closely related to mitochondrial dynamics, has also been suggested as an important factor affecting human response to cold. The present study aimed to clarify the relationship between mitochondrial haplogroup and NST during periods of mild cold exposure.

**Methods:** Seventeen healthy university students (D: n = 8, non-D: n = 9) participated in the present study during summer and winter. A climate chamber was programmed so that ambient temperature inside dropped from 28°C to 16°C over the course of an 80-minute period. Physiological parameters were recorded throughout the course of the experiments.

**Results:** Increases in  $\dot{V}O_2$  were significantly greater during periods of cold exposure in winter than they were during periods of cold exposure in summer, and individuals from the D group exhibited greater winter values of  $\Delta\dot{V}O_2$  than individuals from the non-D group.  $T_{re}$  was significantly lower during periods of rest and cold exposure in winter; however, no significant difference was observed between  $T_{re}$  values of individuals in the D and non-D groups. In addition, although  $\bar{T}_{dist}$  was significantly lower during periods of rest in winter than it was during those same periods in summer, no significant seasonal differences in values of  $\bar{T}_{dist}$  were observed during periods of cold exposure.

**Conclusions:** Results of the present study indicated that NST was greater in winter, and that the D group exhibited greater NST than the non-D group during winter. Despite the differences between groups in NST, no significant differences in rectal and skin temperatures were found between groups in either season. Therefore, it was supposed that mitochondrial DNA haplogroups had a greater effect on variation in energy expenditure involving NST than they had on insulative responses. Future studies are necessary in order to investigate more multiple candidate genes related to human cold adaptation and to elucidate the relationship between gene polymorphism and physiological polytypism.

**Keywords:** mtDNA haplogroup, NST,  $\dot{V}O_2$ , Cold exposure, Seasonal acclimatization

\* Correspondence: takayuki-n@nagasaki-u.ac.jp

<sup>1</sup>Department of Public Health, Nagasaki University Graduate School of Biomedical Sciences, Nagasaki, Japan

Full list of author information is available at the end of the article

## Background

*Homo sapiens* originated in Africa approximately 160,000 year ago, after which time the species rapidly spread around the world during the last glacial period (110,000 to 12,000 years ago) [1]. Our ancestors adapted to various environments during this migration, with both cultural and physiological adaptations proving necessary in order to survive in cold environments [2]. Physiological adaptations to cold, such as the metabolic adaptation of the Inuit [3] and the insulative adaptation of Australian aborigines [4], are well known. Adaptations such as these might have involved genetic adaptations, since these groups were settled in their respective environments for long periods of time. On the other hand, individual variations in both metabolic and insulative type with respect to cold stimuli have also been reported as physiological polymorphism within populations [5].

In urban residents, physiological responses to cold environments include vasoconstriction, which occurs rapidly in response to cold exposure in order to decrease heat loss; however, the range to which the thermal environment can be adjusted by vasoconstriction alone is narrow, and thermogenesis is typically required to maintain optimal body temperature. Thermogenesis can be divided into shivering thermogenesis (ST) and non-shivering thermogenesis (NST). These physiological responses to cold are affected by various environmental or individual factors such as season [6-8], lifestyle [9], and physical characteristics [10]. Recently, NST via metabolism of free fatty acids (FFA) by brown adipose tissue (BAT) has been determined to be an important source of metabolic heat in cold environments [11]. Furthermore, NST activated by BAT was determined to be greater either in winter [12] or after cold acclimatization [13]. Although genetic factors must exist, few studies examining the effects of genetic factors on physiological responses to cold have been undertaken.

In order to examine genetic factors, the present study focused on mitochondria and the mitochondrial genome. Mitochondria are organelles in the cell that generate ATP and heat by OXPHOS (oxidative phosphorylation); this heat is the main source of heat contributing to human body temperature [14]. Furthermore, mitochondria play an important role in energy metabolism involving NST [14,15]. Mitochondria also possess their own genome (mtDNA), with mtDNA polymorphism having been used to help understand the origins of humanity and our moving history [1]. In addition, previous studies have suggested that mtDNA polymorphism was shaped by natural selection, especially in regions of cold climate, and that some haplogroups determined by mtDNA polymorphism had been specifically adapted to cold environments [14,16,17]. Previous studies have also indicated that various mtDNA haplogroups are related to maximum oxygen intake ( $VO_{2max}$ ) [18], athletic performance [19], and metabolic

disease [20]. Results of studies such as these indicated that mtDNA haplogroups were associated with human cold adaptation, and that they affect energy expenditure in particular.

Despite the multitude of studies addressing the role of mtDNA haplogroups in human cold adaptation, no previous studies have directly evaluated cold tolerance. Our previous study aimed to examine the relationship between mtDNA haplogroup and physiological response to serve cold exposure (10°C) [21]. Results indicated that individuals of haplogroup D maintained higher core body temperatures in summer than individuals of haplogroup non-D, although no difference in body temperature could be detected between the two groups in winter. Haplogroup D tended to metabolize greater amounts of fat in winter than did haplogroup non-D, which indicated a greater level of NST in haplogroup D [21]. On the other hand, our previous experiment employed significant levels of cold exposure, which made it difficult to separate ST from NST as all participants shivered in both winter and summer. The present study aimed to clarify the relationship between mtDNA haplogroup and NST by employing periods of mild cold exposure during both summer and winter.

## Method

### Participants

Seventeen healthy university students (Japanese, male, 20 to 24 years old) who exhibited no clinical problems participated in the present study. After having the experimental procedure fully described to them, they consented to their participation in writing. The haplogroups of non-D subjects were M7 (4 participants), F (1 participants), B4 (3 participants), and N9a (1 participants). Table 1 shows the morphological characteristics of the D and non-D groups in each season. Body mass index (BMI) was calculated as follows:

$$BMI = \text{Weight}(\text{kg}) / \text{Height}(\text{m})^2$$

Body surface area (BSA) was calculated using Krazumi's Formula [22]. Experiments were performed with approval from the Ethics Committee of the Graduate School of Design, Kyushu University.

### DNA analysis

Total DNA was extracted from hair shafts by digestion in extraction buffer using ISOHAIR (Code Number 319-03401; Nippon Gene, Tokyo, Japan). The mtDNA spacer D-loop was amplified by PCR using primers M13RV-L15996 and M13(-21)-H408. The analyzed sequences of the D-loop primers were as follows:

mtDNA L15996, 5'-CTCCACCATTTAGCACCCAAA  
GC-3'; and

mtDNA H408, 5'-CTGTTAAAAGTGCATACCGCCA-3'.

**Table 1 Participants' morphological characteristics**

Season	Haplogroup	Height (cm)	Body mass (kg)	BMI	BSA (cm <sup>2</sup> )	Body fat (%)
Summer	D (n = 8)	173.7 ± 7.4	60.8 ± 7.8	19.8 ± 2.2	1.71 ± 0.12	12.3 ± 2.2
	non-D (n = 9)	171.4 ± 8.6	57.6 ± 7.4	19.9 ± 2.0	1.68 ± 0.12	13.1 ± 2.5
Winter	D (n = 8)	173.4 ± 7.2	61.2 ± 6.4	20.3 ± 1.6	1.72 ± 0.11	12.8 ± 2.4
	non-D (n = 9)	170.8 ± 8.9	57.8 ± 7.6	19.8 ± 1.1	1.67 ± 0.12	13.6 ± 2.4

No significant differences in morphological characteristics were found between groups during summer and winter.

The thermocycling profile consisted of an initial de-naturation step at 94°C for 1 minute, followed by 32 cycles of 30 seconds at 94°C, 30 seconds at 56°C, and 75 seconds at 72°C. Purified DNA was sequenced in both directions using an ABI PRISM 310 Genetic Analyzer (Applied Biosystems, Foster City, CA, USA) with a BigDye Terminator v3.1 Cycle Sequencing Kit (Applied Biosystems, Foster City, CA, USA).

### Study procedure

Experiments were conducted in summer (August to September) and winter (February to March) in Fukuoka, Japan. Average temperature in Fukuoka was 28.3°C in summer and 8.5°C in winter. Participants abstained from food and drink for a period of greater than two hours prior to entering the climate chamber. Various measurement sensors (temperatures sensors, gas analyzer) were attached to participants in an environment with a temperature of 28°C prior to experimentation for a period of 30 minutes. After participants had rested quietly for a period of 20 minutes after entering the climate chamber, ambient temperature within the climate chamber dropped from 28°C to 16°C over the course of an approximately 80-minute period. Parameters recorded were rectal temperature, skin temperature (at 7 locations), and oxygen intake during 'rest time' (0 to 20 minutes) and 'cold exposure' (20 to 100 minutes). An electromyogram and a subjective evaluation were also carried out.

Rectal temperature ( $T_{re}$ ) probes were inserted at a depth of 13 cm beyond the anal sphincter. Skin temperature sensors were attached with surgical tape to measurement sites on the forehead, abdomen, forearm, hand, thigh, leg, and foot. Measurements were taken at intervals of 10 seconds using a data logger (LT-8A, Gram Corporation, Saitama, Japan).

Skin temperature was calculated using the seven-point method of Hardy-DuBois [23]. Distal skin temperature ( $\bar{T}_{dist}$ ) was derived using the following equation:

$$\bar{T}_{dist} = (0.14 \times T_{forearm} + 0.05 \times T_{hand} + 0.07 \times T_{foot} + 0.13 \times T_{leg}) / 0.39$$

Oxygen intake ( $VO_2$ ) and carbon dioxide output ( $VCO_2$ ) were measured using a respiratory gas analyzer (AE-300S, Minato Medical Science, Osaka, Japan) through a breathing

tube with a Rudolph mask used to measure expired gas (Rudolph mask, Nihon Kohden, Tokyo, Japan). Respiratory exchange ratio (RER) was calculated as  $VCO_2/VO_2$ . Higher RER values indicated metabolism of glucose and lower RER values indicated metabolism of fat. Changes in the pectoralis major muscle were recorded using an electromyograph (PolyTele, Nihon Santeku, Kyoto, Japan). Data were recorded at a sampling frequency of 1000 Hz and a bandpass filter (20 to 500 Hz) was used in analysis. Electromyographic data obtained during cold exposure was based on muscular changes during the first ten minutes of rest time.

### Statistical analysis

Morphological data were compared by paired *t*-test. Physiological data were compared using three-way (haplogroup, season, and time) analysis of variance (ANOVA). All data were expressed as means ± standard error, and statistical significance was determined at  $P < 0.05$ .

### Results

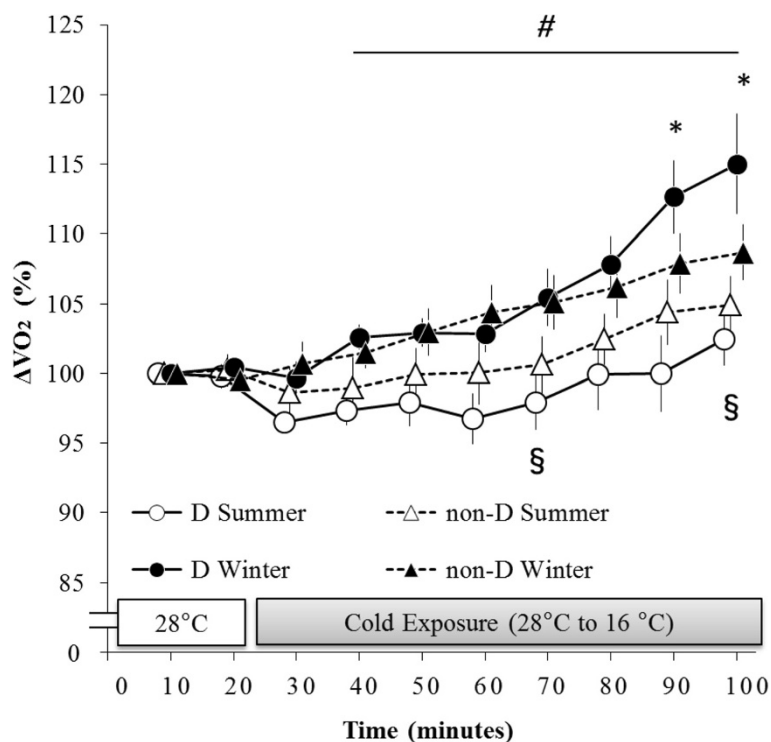
#### Changes in oxygen intake ( $\Delta VO_2$ )

The main effects of season ( $F(1, 15) = 17.07, P < 0.001$ ) and time ( $F(9, 135) = 12.10, P < 0.001$ ) were significant for  $\Delta VO_2$  (Figure 1). There were also significant interactions between season and group ( $F(1, 15) = 5.16, P < 0.001$ ), season and time ( $F(9, 135) = 8.96, P < 0.001$ ), and group, season, and time ( $F(9, 135) = 3.23, P < 0.005$ ).

In a *post-hoc* test carried out in winter,  $\Delta VO_2$  of haplogroup D was significantly greater during the period ranging from 90 to 100 minutes compared with  $\Delta VO_2$  of haplogroup non-D during that same period.  $\Delta VO_2$  of haplogroup D was significantly greater during the period ranging from 40 to 100 minutes in winter than it was during that period in summer.  $\Delta VO_2$  of haplogroup non-D was significantly greater at 70 minutes and 100 minutes in winter than it was at those same points in summer.

#### Change in electromyogram (EMG)

Electromyographic data exhibited no significant main effects for season or time, and no significant interaction was observed between season and time (Figure 2).



**Figure 1 Changes in  $\text{VO}_2$  ( $\Delta\text{VO}_2$ ).** White circles connected by the solid line indicate summer data associated with haplogroup D ( $n = 8$ ) and white triangles connected by the dotted line indicate summer data associated with haplogroup non-D ( $n = 9$ ). Black circles connected by the solid line indicate winter data associated with haplogroup D ( $n = 8$ ) and black triangles connected by the dotted line indicate winter data associated with haplogroup non-D ( $n = 9$ ).  $\Delta\text{VO}_2$  of haplogroup D was significantly higher than that of haplogroup non-D during the period ranging from 90 to 100 minutes in winter.  $\Delta\text{VO}_2$  of haplogroup D was significantly higher during the period ranging from 40 to 100 minutes in winter than it was during that same period in summer.  $\Delta\text{VO}_2$  of haplogroup non-D was significantly higher at both 70 minutes and 100 minutes in winter than it was at those same intervals in summer. \* $P < 0.05$ , comparisons between haplogroups D and non-D in winter. # $P < 0.05$ , comparisons between haplogroup D values taken in summer and winter. § $P < 0.05$ , comparisons between haplogroup non-D values taken in summer and winter.

### Respiratory exchange ratio (RER)

The main effect of season ( $F(1, 15) = 18.22, P < 0.001$ ) was significant for RER (Figure 3). Significant interactions were also detected between season and time ( $F(9, 135) = 5.07, P < 0.001$ ), and group, season, and time ( $F(9, 135) = 2.04, P < 0.05$ ).

In a *post-hoc* test, RER of haplogroup D was significantly lower during the period ranging from 0 to 100 minutes in winter than it was during that same period in summer. RER of haplogroup non-D was significantly lower during the period ranging from 30 to 100 minutes in winter than it was during that same period in summer. Despite these seasonal differences, no significant differences in RER were detected between groups.

### Rectal temperature ( $T_{re}$ )

The main effect of time ( $F(9, 135) = 39.73, P < 0.001$ ) was significant for  $T_{re}$  (Figure 4). A significant interaction was also detected between season and time ( $F(9, 135) = 2.33, P < 0.05$ ). In a *post-hoc* test conducted using both groups,  $T_{re}$  was significantly lower during the period ranging from

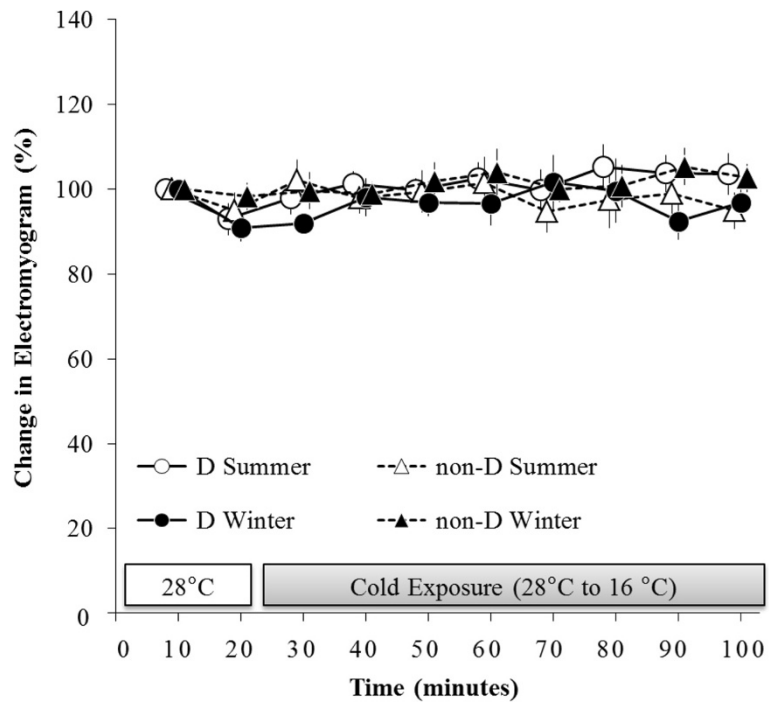
0 to 100 minutes in winter than it was during that same period in summer.

### Distal skin temperature ( $\bar{T}_{dist}$ )

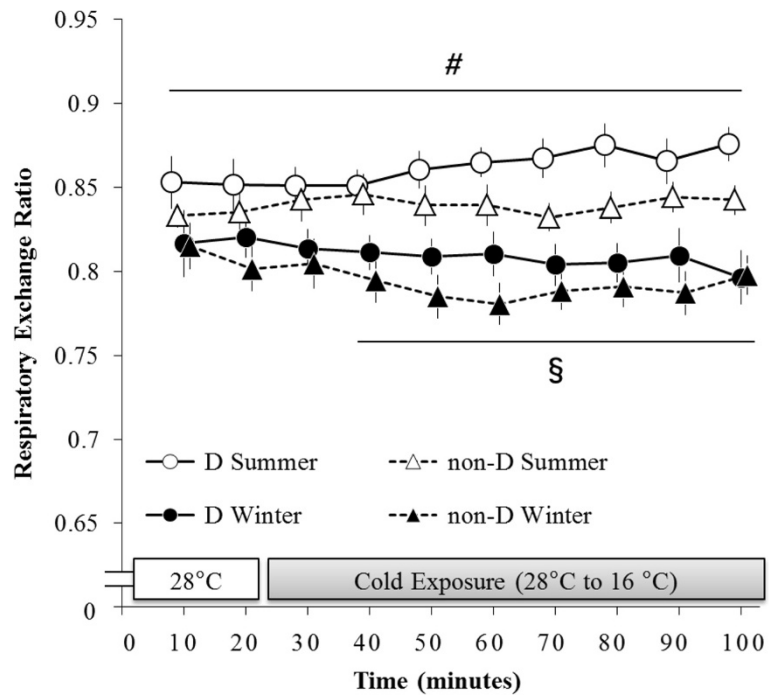
The main effect of time ( $F(9, 135) = 2,049.64, P < 0.001$ ) was significant for  $\bar{T}_{dist}$  (Figure 5). A significant interaction was also detected between season and time ( $F(9, 135) = 29.84, P < 0.001$ ). In a *post-hoc* test conducted using both groups,  $\bar{T}_{dist}$  was significantly lower during the period ranging from 0 to 30 minutes in winter than it was during that same period in summer.

### Discussion

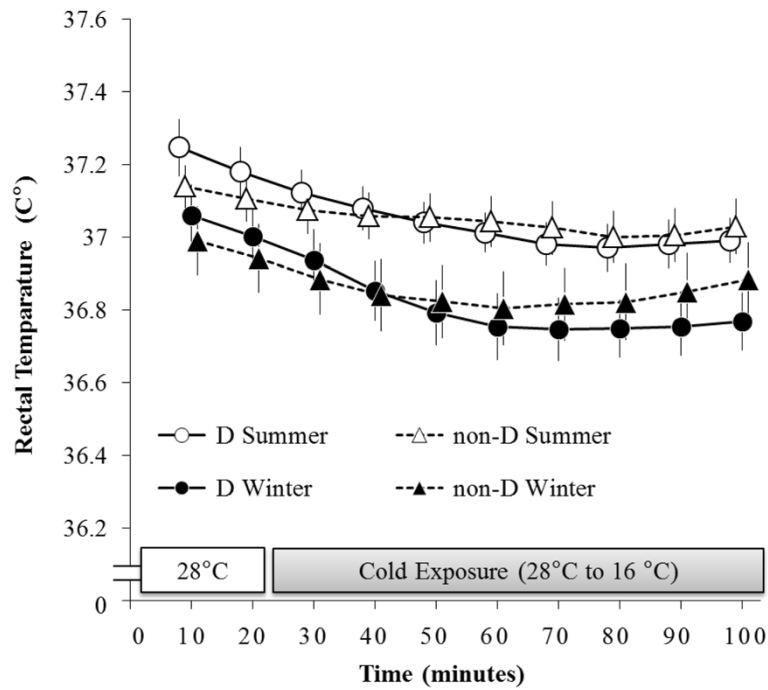
In the present study,  $\Delta\text{VO}_2$  significantly and rapidly increased without shivering in response to cold exposure during winter (Figures 1 and 2). The  $\Delta\text{VO}_2$  of haplogroup D in particular was significantly greater than that of haplogroup non-D in winter. Furthermore, in a result similar to that of our previous study [21], RER of both haplogroups were significantly lower during periods of rest and periods of cold exposure in winter than they were in



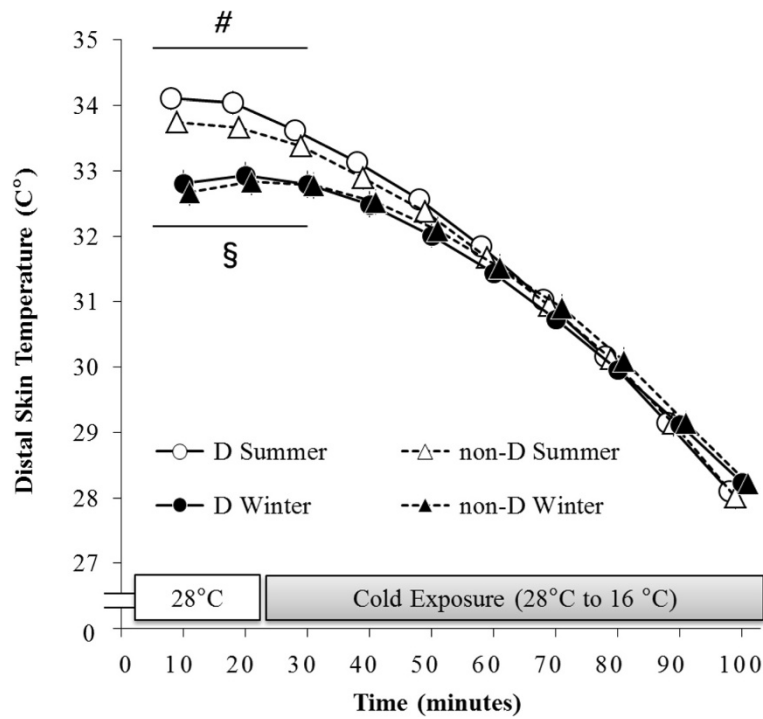
**Figure 2** Changes in electromyogram (EMG). No significant differences existed between season and group.



**Figure 3** Changes in respiratory exchange ratio (RER). RER of haplogroup D was significantly lower during the period ranging from 0 to 100 minutes in winter than it was during that same period in summer. RER of haplogroup non-D was significantly lower during the period ranging from 40 to 100 minutes in winter than it was during that same period in summer. # $P < 0.05$ , comparisons between haplogroup D values taken in summer and winter. § $P < 0.05$ , comparisons between haplogroup non-D values taken in summer and winter.



**Figure 4 Changes in rectal temperature.**  $T_{re}$  of both the D and non-D groups were significantly lower during the period ranging from 0 to 100 minutes in winter than they were during that same period in summer.



**Figure 5 Changes in distal skin temperature ( $\bar{T}_{dist}$ ).**  $\bar{T}_{dist}$  of both the D and non-D groups were significantly lower during the period ranging from 0 to -30 minutes in winter than they were during that same period in summer. # $P < 0.05$ , comparisons between haplogroup D values taken in summer and winter. § $P < 0.05$ , comparisons between haplogroup non-D values taken in summer and winter.



summer (Figure 3). These results suggested that NST stimulated by cold was enhanced in winter, and that haplogroup D exhibited greater NST than did haplogroup non-D in winter.

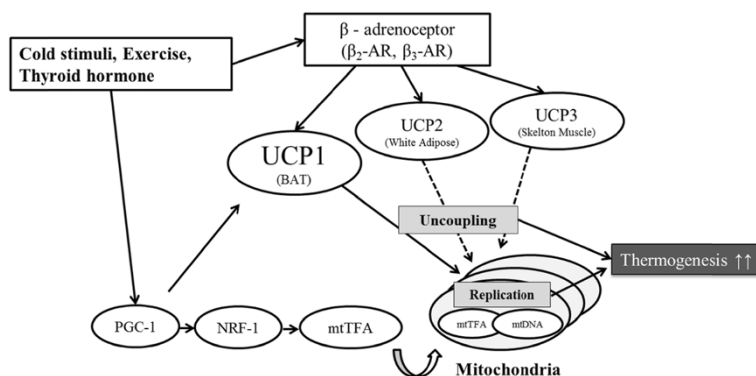
Recent studies suggested that the heat produced by NST is primarily generated by BAT [12,24]. The greater NST of haplogroup D observed in winter might have indicated that the group exhibited greater BAT activity than did haplogroup non-D. Tanaka *et al.* [20] reported that haplogroup D exhibited resistance to metabolic syndrome, which indicated that the greater NST of haplogroup D might have been related to that group's ability to metabolize fat; however, in the present study, no significant difference in fat metabolism as estimated by RER was detected between groups in winter. These results indicated that total heat generated by NST could not be attributed to BAT alone, and that other factors might have contributed to the greater NST observed in haplogroup D.

Previous studies have reported that mtDNA polymorphism affects replication of mitochondria [14]. BAT, as well as skeletal muscle and liver, are mitochondria-rich tissues. Replication of mitochondria is activated by stimuli such as cold, exercise, and thyroid hormone [25]; therefore, the greater NST observed in haplogroup D was likely the result of seasonal cold acclimatization and an increase in mitochondria. BMR (Basal Metabolic Rate) of Japanese individuals has been shown to exhibit seasonal variation, with higher values in winter than in summer [26]. Fat metabolism in Japanese individuals has also been shown to exhibit seasonal variation, being greater in winter than it is in summer [26]. Results such as these have led previous studies to attribute seasonal metabolic changes to changes in mitochondrial function or activity of NST via BAT or other tissues. Although the source of heat generation in NST remains unclear, it can be

concluded that mtDNA haplogroup is a genetic factor that significantly affects  $\Delta VO_2$ .

Despite its impact on  $\Delta VO_2$ , mtDNA haplogroup did not affect  $T_{re}$  (Figure 4) or  $\bar{T}_{dist}$  (Figure 5). Our previous study reported greater values of  $T_{re}$  associated with haplogroup D during periods of severe cold exposure (10°C) in summer; however, decreases in  $T_{re}$  observed in the present study were comparatively lower than those observed in our previous study due to the fact that only mild cold exposure was employed. Therefore, the present study concluded that there existed no significant effect of mtDNA haplogroup on core body temperature. Furthermore, there was no significant effect of haplogroup on  $\bar{T}_{dist}$  in either the present study or our previous study. These results suggested that mitochondrial polymorphism primarily affected thermogenesis involving NST. A similar study employing a more severe or longer cold exposure may allow the effect of mitochondrial polymorphism on body temperature to be elucidated.

One problem existed in that mtDNA haplogroup reflected population structure, and significant differences in  $VO_2$  might have been dependent on other sources of genetic variation. More specifically, differences in NST were affected not only by mitochondrial polymorphism, but also by polymorphisms in other genes, such as *UCP1* and *UCP3*. This dependency was due to the fact that NST is activated by a complex physiological cascade (Figure 6). Hancock *et al.* [27] reported that the genetic diversity of *UCP1* and *UCP3* was shaped by cold climatic conditions, as both genes were related to human energy expenditure involving NST. Polymorphism of *UCP1* affects expression of *UCP1* in BAT [28], and polymorphism of *UCP3* is related to expression of certain proteins in skeletal muscle [29]. Since previous studies suggested that NST was more directly affected by the function of *UCP*,



**Figure 6 Physiological cascade associated with human non-shivering thermogenesis (NST).** NST in humans is activated by a complex physiological cascade. Cold stimuli activate gene expression of *PGC-1alpha* (*Peroxisome proliferator-activated receptor-gamma coactivator-1alpha*), *NRF-1* (*Nuclear respiratory factor 1*), and *mtTFA* (*Mitochondrial transcription factor A*), which in turn promote mitochondrial replication. Due to this process, mitochondrial density increases and NST is enhanced. Cold stimuli also enhance  $\beta_2$ -AR (*beta-2 adrenergic receptor*) and  $\beta_3$ -AR (*beta-3 adrenergic receptor*) via the sympathetic nervous system activating the expression of *UCP1*, *UCP2*, and *UCP3*. This increase in *UCP* also enhances thermogenesis. These flows interact with one other, and each associated gene possesses SNPs that affect physiological function.

mtDNA haplogroup D and other mitochondrial groups might have co-evolved with other gene polymorphisms. These results illustrated the need to develop a more thorough understanding regarding differences in NST, and future studies should focus on the investigation of more gene polymorphisms related to NST.

The present study was limited by its small sample size, which prevented the influence of genetic factors from being excluded, its lack of direct measurement of BAT, and the fact that it did not measure BMR, which is known to affect human cold adaptability. It is also necessary for future studies to investigate more gene polymorphisms related to human cold adaptation, such as *UCPI-3*. Future studies will require greater numbers of participants, measurement of additional physiological parameters, and will need to investigate more gene polymorphisms.

### Perspective

In order to clarify the relationship between physiological polytypism and gene polymorphism with respect to cold adaptation, not only is it necessary to make additional measurements using methodology similar to that of the present study, it is also necessary to develop a new investigational approach. Most importantly, multiple candidate genes related to human cold adaptation should be examined. For example, in the physiological cascade associated with human NST, there exist important genes that either enhance or depress thermogenesis. Candidate genes with the potential to impact cold adaptation or thermogenesis should be narrowed down through population genetics using methodology similar to that of Hancock *et al.* [27]. Investigation into the effects of candidate genes linked to obesity, BMI, or body fat is also necessary, as these phenotypes are believed to be related to energy expenditure. Nakayama *et al.* [30] previously identified *TRIB2* (*tribbles pseudokinase 2*), which is related to visceral fat and obesity. Their results may have also suggested that gene polymorphism of *TRIB2* was shaped by cold climatic conditions in East Asian populations. Nakayama *et al.* [31] also reported that polymorphism of *UCPI* was related to seasonal variation in visceral fat. Research methods such as these are necessary in order to develop an understanding of the relationship between physiological polytypism and gene polymorphism. In order to further advance our understanding of heat generation involving NST, population genetics and field research must identify genes that are potentially linked to human cold adaptation.

Researchers in the field of physiological anthropology should collaborate with researchers in the disciplines mentioned above in order to accumulate physiological data with respect to adaptation. In addition, sample sizes of physiological experiments should be increased to allow for more a robust discussion, and sampling from various regions should be employed to account for variations in the

physical environment such as temperature and day length. In conclusion, due to the importance of genetic research in physiological anthropology, collaboration between population geneticists, field researchers, and physiological researchers will be required in future studies.

### Abbreviations

ANOVA: analysis of variance; BAT: brown adipose tissue; BMI: body mass index; BMR: basal metabolic rate; BSA: body surface area; EMG: electromyogram; FFA: free fatty acids; LCT: lower critical temperature; NST: non-shivering thermogenesis; OXFOS: oxidative phosphorylation; PCR: polymerase chain reaction; RER: respiratory exchange ratio; SNP: single nucleotide polymorphism; ST: shivering thermogenesis;  $\bar{T}_{dist}$ : distal skin temperature;  $T_{re}$ : rectal temperature;  $VCO_2$ : carbon dioxide output;  $VO_2$ : oxygen intake;  $\Delta VO_2$ : changes in  $VO_2$ ;  $VO_{2max}$ : maximum oxygen intake.

### Competing interests

The authors declare that they have no competing interests.

### Authors' contributions

TN carried out the design of the present study and data analysis, and drafted the manuscript. SW contributed to the design of the experiments and checked the manuscript. Both authors read and approved the final manuscript.

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### Author details

<sup>1</sup>Department of Public Health, Nagasaki University Graduate School of Biomedical Sciences, Nagasaki, Japan. <sup>2</sup>Department of Human Science, Faculty of Design, Kyushu University, Fukuoka, Japan.

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### References

1. Relethford JH: Genetic evidence and the modern human origins debate. *Heredity* 2008, **100**:555–563.
2. Steegmann AT: Pearl Memorial Lecture. Human cold adaptation: an unfinished agenda. *Am J Hum Biol* 2007, **22**:218–227.
3. Hart JS, Sabeen HB, Hildes JA, Depocas F, Hammel HT, Andersen KL, Irving L, Foy G: Thermal and metabolic responses of coastal Eskimos during a cold night. *J Appl Physiol* 1962, **17**:953–960.
4. Scholander PF, Hammel HT, Hart JS, Lemessurier DH, Steen J: Cold adaptation in Australian aborigines. *J Appl Physiol* 1958, **13**:211–218.
5. Maeda T: Perspectives on environmental adaptability and physiological polymorphism in thermoregulation. *J Physiol Anthropol Appl Human Sci* 2005, **24**:237–240.
6. Davis TR, Johnston DR: Seasonal acclimatization to cold in man. *J Appl Physiol* 1961, **16**:231–234.
7. Inoue Y, Nakao M, Ueda H, Araki T: Seasonal variation in physiological responses to mild cold air in young and older men. *Int J Biometeorol* 1995, **38**:131–136.
8. Mäkinen TM, Pääkkönen T, Palinkas LA, Rintamäki H, Leppäluoto J, Hassi J: Seasonal changes in thermal responses of urban residents to cold exposure. *Comp Biochem Physiol A Mol Integr Physiol* 2004, **139**:229–238.
9. Maeda T, Sugawara A, Fukushima T, Higuchi S, Ishibashi K: Effects of lifestyle, body composition, and physical fitness on cold tolerance in humans. *J Physiol Anthropol Appl Human Sci* 2005, **24**:439–443.
10. Yasukouchi A, Yamasaki K, Iwanaga K, Fujiwara M, Sato H: Seasonal effects on the relationships between morphological characteristics and decrement of rectal temperature in a cold environment. *Ann Physiol Anthropol* 1983, **2**:39–44. In Japanese with English abstract.
11. van Marken Lichtenbelt WD, Vanhommerig JW, Smulders NM, Drossaerts JM, Kemerink GJ, Bouvy ND, Schrauwen P, Teule GJ: Cold-activated brown adipose tissue in healthy men. *N Engl J Med* 2009, **360**:1500–1508.
12. Saito M, Okamatsu-Ogura Y, Matsushita M, Watanabe K, Yoneshiro T, Nio-Kobayashi J, Iwanaga T, Miyagawa M, Kameya T, Nakada K, Kawai Y, Tsujisaki M: High incidence of metabolically active brown adipose tissue in healthy adult humans: effects of cold exposure and adiposity. *Diabetes* 2009, **58**:1526–1531.



13. van der Lans AA, Hoeks J, Brans B, Vijgen GH, Visser MG, Vosselman MJ, Hansen J, Jörgensen JA, Wu J, Mottaghy FM: **Schrauwen P, van Marken Lichtenbelt WD: Cold acclimation recruits human brown fat and increases nonshivering thermogenesis.** *J Clin Invest* 2013, **123**:3395–3403.
14. Wallace DC: **A mitochondrial paradigm of metabolic and degenerative diseases, aging, and cancer: a dawn for evolutionary medicine.** *Annu Rev Genet* 2005, **39**:359–407.
15. van Marken Lichtenbelt WD, Schrauwen P: **Implications of nonshivering thermogenesis for energy balance regulation in humans.** *Am J Physiol Regul Integr Comp Physiol* 2011, **301**:285–296.
16. Mishmar D, Ruiz-Pesini E, Golik P, Macaulay V, Clark AG, Hosseini S, Brandon M, Easley K, Chen E, Brown MD, Sukernik RI, Olckers A, Wallace DC: **Natural selection shaped regional mtDNA variation in humans.** *Proc Natl Acad Sci U S A* 2003, **100**:171–176.
17. Balloux F, Handley LJ, Jombart T, Liu H, Manica A: **Climate shaped the worldwide distribution of human mitochondrial DNA sequence variation.** *Proc Biol Sci* 2009, **276**:3447–3455.
18. Marcuello A, Martínez-Redondo D, Dahmani Y, Casajús JA, Ruiz-Pesini E, Montoya J, López-Pérez MJ, Díez-Sánchez C: **Human mitochondrial variants influence on oxygen consumption.** *Mitochondrion* 2009, **9**:27–30.
19. Mikami E, Fuku N, Takahashi H, Ohiwa N, Scott RA, Pitsiladis YP, Higuchi M, Kawahara T, Tanaka M: **Mitochondrial haplogroups associated with elite Japanese athlete status.** *Br J Sports Med* 2011, **45**:1179–1183.
20. Tanaka M, Takeyasu T, Fuku N, Li-Jun G, Kurata M: **Mitochondrial genome single nucleotide polymorphisms and their phenotypes in the Japanese.** *Ann N Y Acad Sci* 2004, **1011**:7–20.
21. Nishimura T, Motoi M, Niri Y, Hoshi Y, Kondo R, Watanuki S: **Relationship between seasonal cold acclimatization and mtDNA haplogroup in Japanese.** *J Physiol Anthropol* 2012, **31**:22.
22. Kurazumi Y, Tsuchikawa T, Kakutani K, Torii T, Matsubara N, Horikoshi T: **Evaluation of the conformability of the calculation formula for the body surface area of the human body.** *Jpn J Biometeorol* 2009, **39**:101–106.
23. Hardy JD, DuBois EF: **The technique of measuring radiation and convection.** *J Nutr* 1938, **5**:461–475.
24. Yoneshiro T, Aita S, Matsushita M, Kameya T, Nakada K, Kawai Y, Saito M: **Brown adipose tissue, whole-body energy expenditure, and thermogenesis in healthy adult men.** *Obesity* 2011, **19**:13–16.
25. Irrcher I, Adhihetty PJ, Sheehan T, Joseph AM, Hood DA: **PPARGgamma coactivator-1alpha expression during thyroid hormone- and contractile activity-induced mitochondrial adaptations.** *Am J Physiol Cell Physiol* 2003, **284**:1669–1677.
26. Osiba S: **The seasonal variation of basal metabolism and activity of thyroid gland in man.** *Jpn J Physiol* 1957, **7**:355–365.
27. Hancock AM, Clark VJ, Qian Y, Di Rienzo A: **Population genetic analysis of the uncoupling proteins supports a role for UCP3 in human cold resistance.** *Mol Biol Evol* 2011, **28**:601–614.
28. Su AI, Wiltshire T, Batalov S, Lapp H, Ching KA, Block D, Zhang J, Soden R, Hayakawa M, Kreiman G, Cooke MP, Walker JR, Hogenesch JB: **A gene atlas of the mouse and human protein-encoding transcriptomes.** *Proc Natl Acad Sci U S A* 2004, **101**:6062–6067.
29. Schrauwen P, Xia J, Bogardus C, Pratley RE, Ravussin E: **Skeletal muscle uncoupling protein 3 expression is a determinant of energy expenditure in Pima Indians.** *Diabetes* 1999, **48**:146–149.
30. Nakayama K, Ogawa A, Miyashita H, Tabara Y, Igase M, Kohara K, Miki T, Kagawa Y, Yanagisawa Y, Katashima M, Onda T, Okada K, Fukushima S, Iwamoto S: **Positive natural selection of TRIB2, a novel gene that influences visceral fat accumulation, in East Asia.** *Hum Genet* 2013, **132**:201–217.
31. Nakayama K, Miyashita H, Yanagisawa Y, Iwamoto S: **Seasonal effects of UCP1 gene polymorphism on visceral fat accumulation in Japanese adults.** *PLoS One* 2013, **25**:e74720.

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