

ORIGINAL ARTICLE

Open Access

# Seasonal effects of the *UCP3* and the *RPTOR* gene polymorphisms on obesity traits in Japanese adults

Kazuhiro Nakayama<sup>1\*</sup>, Hiroshi Miyashita<sup>2</sup> and Sadahiko Iwamoto<sup>1</sup>

## Abstract

**Background:** Non-shivering thermogenesis (NST) involves a substantial amount of energy expenditure in humans and, thus, contributes to reducing the risk for obesity. Molecular evolutionary studies have reported that SNPs in/near the uncoupling protein 3 gene (*UCP3*) and the regulatory associated protein of mTOR complex 1 gene (*RPTOR*) might influence NST and confer adaptive advantages for modern human dispersal into cold environments. In the present study, the impact of these SNPs on obesity-related traits was investigated.

**Methods:** Study subjects consisted of 2,834 Japanese adults (percentage of female: 46%, mean age: 51.5). Associations of the *UCP3*-55C/T and the *RPTOR*-26934C/T - the 2 potential genetic variations involved in cold adaptation and thermogenic mechanisms in mammals, with quantitative obesity-related traits including body mass index (BMI), waist circumference, visceral fat area (VFA), VFA adjusted for BMI, and selected blood parameters - were tested using multiple linear regression models. Sliding window sampling analysis was applied to depict seasonal effects of the SNPs on the obesity-related phenotypes.

**Results:** *UCP3*-55C/T and the *RPTOR*-26934C/T did not show any association with obesity traits and blood chemical parameters in multiple linear regression models consisting of the whole subjects. Moreover, sliding window sampling-based association analyses involving seasonality also failed to find associations between these two SNPs and obesity-related traits.

**Conclusions:** *UCP3*-55C/T and the *RPTOR*-26934C/T may only have subtle effects on the development of obesity-related traits in the present humans. These two SNPs might be irrelevant to inter-individual variations in energy metabolism and efficiency of NST.

**Keywords:** Uncoupling protein 3 gene, Single nucleotide polymorphism, Visceral fat accumulation, Seasonality

## Background

The principal cause of obesity in humans is the imbalance of energy intake and expenditure. Non-shivering thermogenesis (NST) currently draws much attention as a factor for reducing the risk of obesity through enhancement of energy expenditure. NST mainly occurs in thermogenic adipocytes including brown and beige/brite adipocytes (BAT) [1-4]. The amount of active BAT observed in the human body is strongly influenced by the age of the subject. The age-related decline of BAT is thought to increase risks for obesity in adults [5]. Additionally, activation of BAT has frequently been observed in experiments performed in cold seasons compared to hot seasons [2]. The

seasonality of BAT activity would partly explain the enhancement of energy expenditure in winter [6,7]. NST likely had an adaptive significance in the modern human dispersal into cold environments. The susceptibility to obesity in the present human populations is thus considered to have been partly shaped by past genetic adaptation to cold environments [8,9].

A SNP near the uncoupling protein 1 gene (*UCP1*), namely *UCP1*-3826G/A, is a promising genetic variation linking cold adaptation and resistance to obesity. *UCP1* is a mitochondrial protein highly expressed in BAT and dissipates the energy for ATP synthesis as heat. [10]. The A allele of the -3826 site, which increased the *UCP1* expression levels, was linked with higher BAT activity and reduced risks of obesity [10-13]. A worldwide survey of human genome variations revealed that the A allele

\* Correspondence: nakayama@jichi.ac.jp

<sup>1</sup>Division of Human Genetics, Center for Molecular Medicine, Jichi Medical University, Shimotsuke-shi, Tochigi 329-0498, Japan

Full list of author information is available at the end of the article

was more prevalent in ethnic groups inhabiting higher latitudes. The cline was considered to be shaped by past adaptations to cold environments: the A allele was more adaptive at higher latitudes due to its enhanced activity in NST [14].

Besides the *UCP1*-3826G/A, genetic variations supporting the relationship between past cold adaptation and the present susceptibility to obesity have not been extensively studied [9]. Evolutionary genetic studies have reported several SNPs that showed striking correlation between allele frequencies and geographic or climatic factors [15-17]. Among them, SNPs near uncoupling protein 3 gene (*UCP3*-55C/T) and the regulatory associated protein of the mTOR complex 1 gene (*RPTOR*-26934C/T) were considered to be potential adaptive genetic variation candidates, since molecular and animal studies have suggested relevance of these 2 genes in thermogenic mechanisms in mammals. *UCP3* is a member of the uncoupling protein gene family and possibly takes part in the NST in skeletal muscles [18]. *RPTOR* encodes an intracellular signaling protein that participates in regulation of cell growth [19]. Adipocyte-specific knockout of *Rptor* (mouse homologue of *RPTOR*) induced enhanced mitochondrial respiration and lean phenotypes in mice [20]. These SNPs, therefore, may contribute to susceptibility to obesity similar to *UCP1*-3826G/A. In the present study, we performed genetic associations using a large-scale cohort to assess the impact of the *UCP3* and *RPTOR* SNPs on obesity-related traits in modern humans.

## Methods

### Subjects

The study population comprised 3,013 Japanese adults who attended general health checkups in the Jichi Medical University Healthcare Center during January 2009 to March 2011[8,13]. All participants provided written informed consent. Medical information from the health checkup and genome DNA were obtained from all participants. Visceral fat area (VFA) was measured with the bioelectrical impedance method at the umbilical level [21]. Waist circumference (WC) at the umbilical level was measured using a tape measure. Blood medical traits including fasting plasma glucose (FPG), hemoglobin (Hb) A<sub>1c</sub>, plasma triglycerides (TG), total cholesterol (TC), high density lipoprotein cholesterol (HDL-c) were directly measured using standard methods. Low density lipoprotein cholesterol (LDL-c) levels were estimated using the Friedewald's formula. Individuals who had undergone abdominal surgery were excluded from the analyses. The study design was approved by the Ethical Committee of Jichi Medical University.

### Genotyping

Genotypes of *UCP3*-55C/T (rs1800849) and *RPTOR*-26934C/T (rs11868112) from each participant were determined using

TaqMan SNP Genotyping Assays on a ViiA 7 Real Time PCR System (Thermo Fisher Scientific, Waltham, MA, USA), and KAPA PROBE FAST qPCR kit (Kapa Biosystems, Boston, MA, USA).

### Association analysis

$\chi^2$  goodness-of-fit test was used to assess the Hardy-Weinberg equilibrium of the genotype distribution. In the association analyses, an additive model was assumed. Association of SNP genotypes and quantitative traits was assessed using multiple linear regression models [13]. For body mass index (BMI), WC, and VFA, the number of T alleles (0, 1, and 2), sex, age, speed of walking (based on the health checkup questionnaire question: 'Do you walk faster than other people of same sex and about same ages?') of each participant [13] were included as explanatory variables. Associations of SNPs and obesity-related blood parameters including FPG, HbA<sub>1c</sub>, TG, TC, HDL-c and LDL-c were also tested. In addition to genotypes, sex, age, and BMI were also included as explanatory variables for the blood parameters. Seasonality of SNP effects on VFA was evaluated using the sliding window sampling, which was previously used to show the seasonal effect of *UCP1*-3826G/A on VFA. Briefly, associations of the SNPs with VFA were tested in 12 subsets of the populations, each consisting of individuals sampled during overlapping continuous 3-month periods. Monthly outdoor temperature data measured at the nearest observatory were obtained from the Japan Meteorological Agency. The above-described statistical tests were performed using SPSS version 22 (IBM Corporation, Armonk, NY, USA).

**Table 1 Characteristics of successfully genotyped individuals**

Traits	Male	Female
Numbers of individuals	1,533	1,301
Age	52.5 (9.3)	50.5 (8.6)
VFA (cm <sup>2</sup> )	114.2 (41.7)	62.5 (27.1)
WC (cm)	86.9 (8.5)	80.9 (9.3)
BMI (kg/m <sup>2</sup> )	24.3 (3.2)	22.7 (3.6)
FPG (mg/dl)	104.9 (20.1)	95.1 (13.2)
HbA <sub>1c</sub> (%)	5.4 (0.6)	5.2 (0.5)
TG (mg/dl)	132.7 (88.3)	87.6 (47.6)
TC (mg/dl)	205.8 (32.0)	212.7 (34.8)
HDL-c (mg/dl)	57.7 (13.7)	70.6 (16.3)
LDL-c (mg/dl)	122.5 (28.3)	124.6 (30.7)

Except for the numbers of individuals, mean values (standard deviations) are shown.

BMI, body mass index; FPG, fasting plasma glucose, HbA<sub>1c</sub>, hemoglobin A<sub>1c</sub>; HDL-c, high density lipoprotein cholesterol; LDL-c, low density lipoprotein cholesterol; TC, total cholesterol; TG, triglycerides; VFA, visceral fat area; WA, waist circumference.

**Table 2 Association between obesity-related traits and the tested SNPs**

	CC		CT		TT		P- values
	Mean	SE	Mean	SE	Mean	SE	
<i>UCP3-55C/T</i>							
BMI (kg/m <sup>2</sup> )	23.60	0.09	23.50	0.10	23.52	0.20	>0.05
WC (cm)	84.24	0.24	84.08	0.26	84.13	0.54	>0.05
VFA (cm <sup>2</sup> )	90.60	0.90	90.48	0.95	90.01	2.00	>0.05
aVFA (cm <sup>2</sup> )	90.30	0.52	90.91	0.55	90.43	1.16	>0.05
<i>RPTOR-26934C/T</i>							
BMI (kg/m <sup>2</sup> )	23.47	0.10	23.60	0.10	23.60	0.16	>0.05
WC (cm)	84.21	0.26	84.16	0.25	84.06	0.43	> 0.05
VFA (cm <sup>2</sup> )	90.90	0.97	90.37	0.93	90.09	1.61	> 0.05
aVFA (cm <sup>2</sup> )	91.36	0.56	90.11	0.54	89.62	0.93	> 0.05

Mean values and the standard errors (SE) in each genotype group (CC, CT, and TT) are indicated. These values were adjusted for sex, age, age<sup>2</sup>, and walking speed. P-values of T allele copy numbers (0, 1, and 2) in multiple linear regression models are indicated.

aVFA, VFA adjusted for BMI; BMI, body mass index; VFA, visceral fat area; WA, waist circumference.

### Results and Discussion

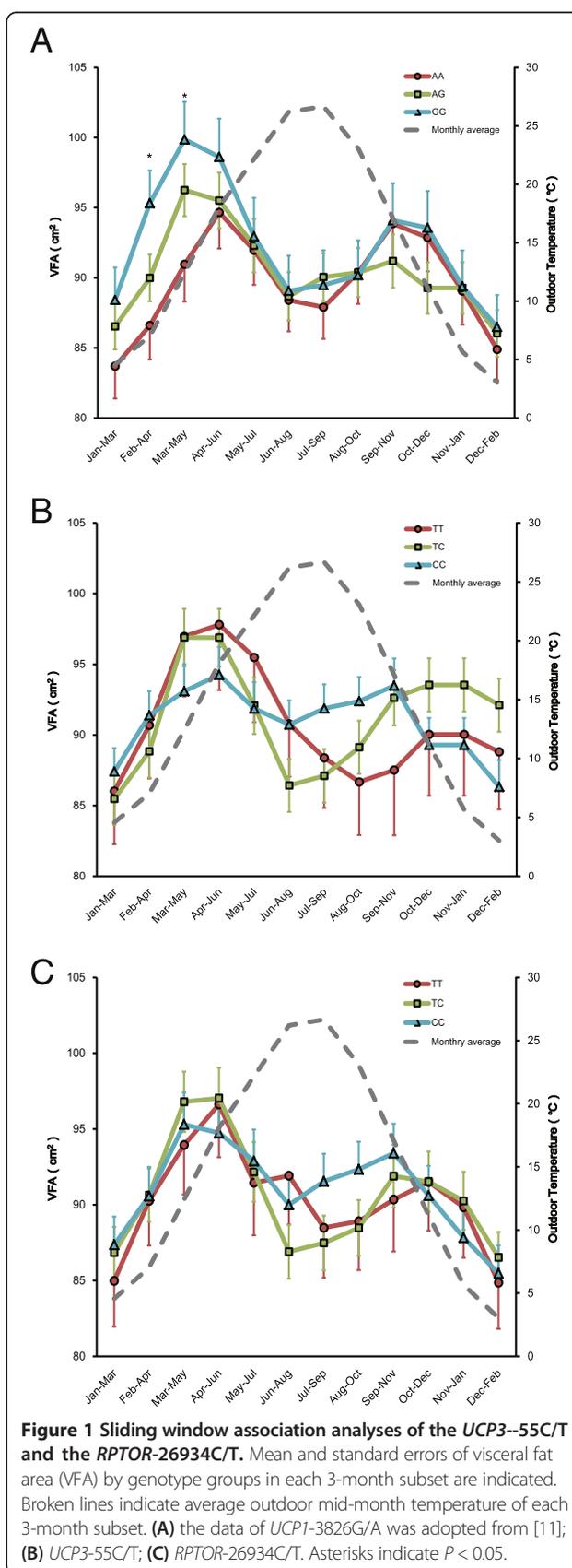
In total, 2,834 successfully genotyped individuals, who had not undergone abdominal surgery, were included for further analyses. The characteristics of the genotyped subjects are summarized in Table 1. The genotype distribution of *UCP3-55C/T* was as follow: CC = 1,356, CT = 1,205, and TT = 273. No deviation from the Hardy-Weinberg equilibrium was observed for the genotype distribution ( $P = 0.824$ ). *RPTOR-26934C/T* genotypes were as follow: CC = 1,158, CT = 1,262, and TT = 414. Although a slight excess of homozygotes was observed ( $P = 0.021$ ), genotype and allele frequencies of *RPTOR-26934C/T* were comparable to the previously reported values. No significant differences in male/female proportion and age distribution were observed among *UCP3-55C/T* genotype groups ( $P = 0.823$ ,  $\chi^2$  test). The *RPTOR-26934C/T* showed slight sex differences in genotype distribution ( $P = 0.024$ ,  $\chi^2$  test).

**Table 3 Associations between the tested SNPs and selected blood parameters**

Traits	<i>UCP3-55C/T</i>		<i>RPTOR-26934C/T</i>	
	Effects	P-values	Effects	P-values
FPG	-0.013	0.431	0.001	0.974
HbA <sub>1c</sub>	-0.022	0.210	-0.007	0.678
TG	0.005	0.779	-0.017	0.294
TC	0.030	0.105	0.015	0.410
HDL-c	-0.003	0.834	0.027	0.100
LDL-c	0.032	0.080	0.004	0.847

Effects ( $\beta$  coefficients) and P-values of T allele copy numbers (0, 1, and 2) in multiple linear regression models are indicated.

FPG, fasting plasma glucose, HbA<sub>1c</sub>, hemoglobin A<sub>1c</sub>; HDL-c, high density lipoprotein cholesterol; LDL-c, low density lipoprotein cholesterol; TC, total cholesterol; TG, triglycerides.



Age distribution was not different between genotypes ( $P > 0.05$ , 1-way analysis of variance).

For both of *UCP3-55C/T* and *RPTOR-26934C/T*, the T allele was supposed to be linked with higher thermogenic activity since the allele frequency of the T allele was shown to be higher in ethnic groups at higher latitudes [14,17]. Thus, the effects of T allele copy number (0, 1, or 2) on obesity-related quantitative traits were tested in each subject using multiple linear regression models (Table 2). We first tested the models comprising whole individuals. Mean values of BMI, WC, VFA and VFA adjusted for BMI were similar among genotype groups of both SNPs and no significant effect of the T allele copy number on the obesity-related traits was observed. Additionally, these two SNPs did not show associations with blood medical parameters including FPG, HbA<sub>1c</sub>, TG, TC, HDL-c and LDL-c (Table 3).

A previous study had revealed that the stratification of subjects by season of examination successfully refined the association between the *UCP1-3826G/A* and VFA [13]. This phenomenon was likely due to the physiological nature of BAT and VFA, wherein cold stress was required to enhance the NST, and VFA reflected the energy balance more sensitively than BMI or WC [11,13]. If the *UCP3* and the *RPTOR* contributed to the efficiency of NST, a similar analysis would be useful for these 2 SNPs. Therefore, we assessed the seasonal effects of these two SNPs on the VFA using the sliding window sampling method. For the *UCP1-3826G/A*, carriers of the A allele showed smaller mean VFA during winter to spring, and the genotypic differences in VFA were significant in subsets sampled during February to May (Figure 1A, adopted from [13]). For *UCP3-55C/T* and *RPTOR-26934C/T* (Figure 1B and 1C), the mean values of VFA were very similar in these subsets ( $P > 0.10$ ). Both SNPs showed a similar trend where the T allele showed smaller VFA during summer to autumn, but the associations were not significant ( $P > 0.098$ ).

There are certain limitations to our study. Our large-scale cohort allowed us to test seasonality of the effect of SNPs; however, longitudinal analyses, instead of the sliding window analysis, may be more preferable to depict seasonal effect. Moreover, precise information about physical activity and dietary intakes, which should be controlled in the association analyses of obesity-related traits, was unavailable in our cohort. The present multiple linear regression models were not adjusted for skeletal muscle mass, which would have a large impact on obesity-related traits [22,23]. Moreover, winter temperature of the study site (Kanto, Japan) was possibly not low enough to stimulate NST in skeletal muscle. Considering the nature of NST in skeletal muscle might be required to test the association of the *UCP3* and obesity-related traits. The present study tested a single SNP by a locus, but other SNPs influencing obesity-related traits have been reported for the *UCP3*

and the *RPTOR* [24-27]. Therefore, the nearby SNPs and their haplotypes should be assessed for their association with obesity-related traits. Relevance of these two SNPs to NST must be tested by precise physiological genetic experiments using a climate chamber [28].

## Conclusions

Overall, our genetic association analyses including seasonality on a large-scale cohort showed that the *UCP3-55C/T* and the *RPTOR-26934C/T* did not influence obesity-related traits. Roles of the *UCP3* and the *RPTOR* in NST remain unclear but the effects of these two SNPs on energy metabolism balance and, perhaps, on efficiency of NST in humans may be subtle. Climate-associated natural selection on the *UCP3-55C/T* and the *RPTOR-26934C/T* can be explained by physiological functions other than NST. *UCP3* has been shown to be involved in the regulation of reactive oxygen species in mitochondria and, thus, may play a role in the maintenance of mitochondrial functions [29]. *RPTOR* encodes a subunit of the mammalian target of rapamycin (mTOR) complex 1 (mTORC1), which is a protein complex regulating protein synthesis in response to nutrition, energy, and cellular redox status [19]. The mTORC1 participates in various biological pathway including cell proliferation, angiogenesis, and various immune systems. Adaptation to pathogens, for instance, may explain the climate associated evolution of *RPTOR-26934C/T* [17]. Associations of these two SNPs with other various physiological traits should be tested in future. Finally, the signature of adaptive evolution for the *UCP3* and the *RPTOR* should be re-verified using molecular evolutionary methods other than those adopted by Hancock *et al.* and Sun *et al.* [14-17].

## Abbreviations

BAT: brown and beige/brite adipose tissues; BMI: body mass index; FPG: fasting plasma glucose; Hb: hemoglobin; HDL-c: high density lipoprotein cholesterol; LDL-c: low density lipoprotein cholesterol; mTORC1: mammalian target of rapamycin complex 1; NST: non-shivering thermogenesis; PCR: polymerase chain reaction; RPTOR: regulatory associated protein of mTOR complex 1; SNP: single nucleotide polymorphism; TC: total cholesterol; TG: triglycerides; UCP1: uncoupling protein 1; UCP3: uncoupling protein 3; VFA: visceral fat area; WC: waist circumference.

## Competing interests

The authors declare that they have no competing interests.

## Authors' contributions

KN designed the study, carried out the experiments, analyzed data, and drafted the paper. HM and SI participated in the study design and managed the sample correction. All authors read and approved the final manuscript.

## Acknowledgements

We are grateful to the staff of the Jichi Medical University Health Care Center. This study was supported in part by MEXT KAKENHI: Grant Numbers 25291103, 26291096 (KN).

## Author details

<sup>1</sup>Division of Human Genetics, Center for Molecular Medicine, Jichi Medical University, Shimotsuke-shi, Tochigi 329-0498, Japan. <sup>2</sup>Jichi Medical University Health Care Center, Shimotsuke-shi, Tochigi 329-0434, Japan.

Received: 1 October 2014 Accepted: 2 December 2014  
Published: 22 December 2014

## References

1. Cypess AM, Lehman S, Williams G, Tal I, Rodman D, Goldfine AB, Kuo FC, Palmer EL, Tseng YH, Doria A, Kolodny GM, Kahn CR: **Identification and importance of brown adipose tissue in adult humans.** *N Engl J Med* 2009, **360**(15):1509–1517.
2. Saito M, Okamatsu-Ogura Y, Matsushita M, Watanabe K, Yoneshiro T, Nio-Kobayashi J, Iwanaga T, Miyagawa M, Kameya T, Nakada K, Kawai Y, Tsujisaki M: **High incidence of metabolically active brown adipose tissue in healthy adult humans: effects of cold exposure and adiposity.** *Diabetes* 2009, **58**(7):1526–1531.
3. van Marken Lichtenbelt WD, Vanhommerig JW, Smulders NM, Drossaerts JM, Kemerink GJ, Bouvy ND, Schrauwen P, Teule GJ: **Cold-activated brown adipose tissue in healthy men.** *N Engl J Med* 2009, **360**(15):1500–1508.
4. Virtanen KA, Lidell ME, Orava J, Heglind M, Westergren R, Niemi T, Taittonen M, Laine J, Savisto NJ, Enerback S, Nuutila P: **Functional brown adipose tissue in healthy adults.** *N Engl J Med* 2009, **360**(15):1518–1525.
5. Yoneshiro T, Aita S, Matsushita M, Okamatsu-Ogura Y, Kameya T, Kawai Y, Miyagawa M, Tsujisaki M, Saito M: **Age-related decrease in cold-activated brown adipose tissue and accumulation of body fat in healthy humans.** *Obesity* 2011, **19**(9):1755–1760.
6. van Ooijen AM, van Marken Lichtenbelt WD, van Steenhoven AA, Westerterp KR: **Seasonal changes in metabolic and temperature responses to cold air in humans.** *Physiol Behav* 2004, **82**(2–3):545–553.
7. Mäkinen TM, Pääkkönen T, Palinkas LA, Rintamäki H, Leppäluoto J, Hassi J: **Seasonal changes in thermal responses of urban residents to cold exposure.** *Comp Biochem Physiol A Mol Integr Physiol* 2004, **139**(2):229–238.
8. Nakayama K, Ogawa A, Miyashita H, Tabara Y, Igase M, Kohara K, Miki T, Kagawa Y, Yanagisawa Y, Katashima M, Onda T, Okada K, Fukushima S, Iwamoto S: **Positive natural selection of TRIB2, a novel gene that influences visceral fat accumulation, in East Asia.** *Hum Genet* 2013, **132**(2):201–217.
9. Sellayah D, Cagampang FR, Cox RD: **On the evolutionary origins of obesity: a new hypothesis.** *Endocrinology* 2014, **155**(5):1573–1588.
10. Rose G, Crocco P, D'Aquila P, Montesanto A, Bellizzi D, Passarino G: **Two variants located in the upstream enhancer region of human UCP1 gene affect gene expression and are correlated with human longevity.** *Exp Gerontol* 2011, **46**(11):897–904.
11. Yoneshiro T, Ogawa T, Okamoto N, Matsushita M, Aita S, Kameya T, Kawai Y, Iwanaga T, Saito M: **Impact of UCP1 and beta3AR gene polymorphisms on age-related changes in brown adipose tissue and adiposity in humans.** *Int J Obes (Lond)* 2013, **37**(7):993–998.
12. Nagai N, Sakane N, Tsuzaki K, Moritani T: **UCP1 genetic polymorphism (–3826 A/G) diminishes resting energy expenditure and thermoregulatory sympathetic nervous system activity in young females.** *Int J Obes (Lond)* 2011, **35**(8):1050–1055.
13. Nakayama K, Miyashita H, Yanagisawa Y, Iwamoto S: **Seasonal effects of UCP1 gene polymorphism on visceral fat accumulation in Japanese adults.** *PLoS One* 2013, **8**(9):e74720.
14. Hancock AM, Clark VJ, Qian Y, Di Rienzo A: **Population genetic analysis of the uncoupling proteins supports a role for UCP3 in human cold resistance.** *Mol Biol Evol* 2011, **28**(1):601–614.
15. Hancock AM, Witonsky DB, Gordon AS, Eshel G, Pritchard JK, Coop G, Di Rienzo A: **Adaptations to climate in candidate genes for common metabolic disorders.** *PLoS Genet* 2008, **4**(2):e32.
16. Hancock AM, Witonsky DB, Alkorta-Aranburu G, Beall CM, Gebremedhin A, Sukernik R, Utermann G, Pritchard JK, Coop G, Di Rienzo A: **Adaptations to climate-mediated selective pressures in humans.** *PLoS Genet* 2011, **7**(4):e1001375.
17. Sun C, Southard C, Witonsky DB, Kittler R, Di Rienzo A: **Allele-specific down-regulation of RPTOR expression induced by retinoids contributes to climate adaptations.** *PLoS Genet* 2010, **6**(10):e1001178.
18. Brand MD, Esteves TC: **Physiological functions of the mitochondrial uncoupling proteins UCP2 and UCP3.** *Cell Metab* 2005, **2**(2):85–93.
19. Hara K, Maruki Y, Long X, Yoshino K, Oshiro N, Hidayat S, Tokunaga C, Avruch J, Yonezawa K: **Raptor, a binding partner of target of rapamycin (TOR), mediates TOR action.** *Cell* 2002, **110**(2):177–189.
20. Polak P, Cybulski N, Feige JN, Auwerx J, Ruegg MA, Hall MN: **Adipose-specific knockout of raptor results in lean mice with enhanced mitochondrial respiration.** *Cell Metab* 2008, **8**(5):399–410.
21. Ryo M, Maeda K, Onda T, Katashima M, Okumiya A, Nishida M, Yamaguchi T, Funahashi T, Matsuzawa Y, Nakamura T, Shimomura I: **A new simple method for the measurement of visceral fat accumulation by bioelectrical impedance.** *Diabetes Care* 2005, **28**(2):451–453.
22. Gallagher D, Visser M, De Meersman RE, Sepúlveda D, Baumgartner RN, Pierson RN, Harris T, Heymsfield SB: **Appendicular skeletal muscle mass: effects of age, gender, and ethnicity.** *J Appl Physiol* 1997, **83**(1):229–239.
23. Abe T, Kearns CF, Fukunaga T: **Sex differences in whole body skeletal muscle mass measured by magnetic resonance imaging and its distribution in young Japanese adults.** *Br J Sports Med* 2003, **37**(5):436–440.
24. Berndt SI, Gustafsson S, Magi R, Ganna A, Wheeler E, Feitosa MF, Justice AE, Monda KL, Croteau-Chonka DC, Day FR, Esko T, Fall T, Ferreira T, Gentilini D, Jackson AU, Luan J, Randall JC, Vedantam S, Willer CJ, Winkler TW, Wood AR, Workalemahu T, Hu YJ, Lee SH, Liang L, Lin DY, Min JL, Neale BM, Thorleifsson G, Yang J, et al: **Genome-wide meta-analysis identifies 11 new loci for anthropometric traits and provides insights into genetic architecture.** *Nat Genet* 2013, **45**(5):501–512.
25. Qian L, Xu K, Xu X, Gu R, Liu X, Shan S, Yang T: **UCP2–866G/A, Ala55Val and UCP3–55C/T polymorphisms in association with obesity susceptibility - A meta-analysis study.** *PLoS One* 2013, **8**(4):e58939.
26. de Souza BM, Brondani LA, Boucas AP, Sortica DA, Kramer CK, Canani LH, Leitao CB, Crispim D: **Associations between UCP1–3826A/G, UCP2–866G/A, Ala55Val and Ins/Del, and UCP3–55C/T polymorphisms and susceptibility to type 2 diabetes mellitus: case-control study and meta-analysis.** *PLoS One* 2013, **8**(1):e54259.
27. Brondani A, Assmann TS, de Souza BM, Boucas AP, Canani LH, Crispim D: **Meta-analysis reveals the association of common variants in the uncoupling protein (UCP) 1–3 genes with body mass index variability.** *PLoS One* 2014, **9**(5):e96411.
28. Nishimura T, Motoi M, Niri Y, Hoshi Y, Kondo R, Watanuki S: **Relationship between seasonal cold acclimatization and mtDNA haplogroup in Japanese.** *J Physiol Anthropol* 2012, **31**:22.
29. Mailloux RJ, Harper ME: **Mitochondrial proticity and ROS signaling: lessons from the uncoupling proteins.** *Trends Endocrinol Metab* 2012, **23**(9):451–458.

doi:10.1186/1880-6805-33-38

Cite this article as: Nakayama et al.: Seasonal effects of the UCP3 and the RPTOR gene polymorphisms on obesity traits in Japanese adults. *Journal of Physiological Anthropology* 2014 33:38.

Submit your next manuscript to BioMed Central  
and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
www.biomedcentral.com/submit

